



Direct Assembly of Modified Proteins on Carbon Nanotubes in an Aqueous Solution

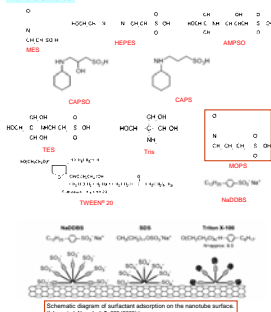
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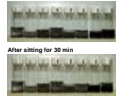
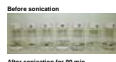
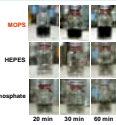
ABSTRACT

Carbon nanotubes (CNTs) have superior mechanical and electrical properties that have opened up many potential applications. However, poor dispersibility and solubility, due to the substantial van der Waals attraction between tubes, have prevented the use of CNTs in practical applications, especially biotechnology applications. Effective dispersion of CNTs into small bundles or individual tubes in solvents is crucial to ensure homogeneous properties and enable practical applications. In addition to dispersion of CNTs into a solvent, the selection of appropriate solvent, which is compatible with a desired matrix, is an important factor to improve the mechanical, thermal, optical, and electrical properties of CNT-based fibers and composites. In particular, dispersion of CNTs into an aqueous system has been a challenge due to the hydrophobic nature of CNTs. Here we show an effective method for dispersion of both single wall CNTs (SWCNTs) and few wall CNTs (FWCNTs) in an aqueous buffer solution. We also show an assembly of cationized P-coated ferritins on the well dispersed CNTs in an aqueous buffer solution.

Buffers Studied



MES: 2-(N-ethylmorpholino)ethanesulfonic acid
HEPES: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
AMPBSO: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
CAPBSO: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
CAPS: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
TES: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
MOPS: 3-(methylmorpholino)propanesulfonic acid
NaOBS: N-(3-(3-dimethylammonio)propyl)carbamoyl-L-homoserine
Tris-HCl: 2-amino-2-hydroxypropane-1,3-diol



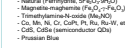
Photos of initial HEPES SWCNT dispersion in the various buffers. MOPS (0.05 M) (1.0 wt. %) with 0.025 M NaCl (0.15 wt. %) pH 7.5 solution containing SWCNT at 0.125 mg/mL HEPES (0.025 M) (0.8 wt. %) with 0.025 M NaCl (0.15 wt. %) pH 7.5 solution containing SWCNT at 0.125 mg/mL and phosphate (0.025 M) (0.8 wt. %) with 0.025 M NaCl (0.15 wt. %) pH 7.2 containing SWCNT at 0.05 mg/mL. All samples were prepared after sonication for 20, 30, and 60 min. Third pictures at each solution were taken after 30 min sonication and then adding 2 ml buffer more with 30 min sonication.

Photos of initial SWCNT (0.5 mg/mL) buffer suspensions after sonication for 30 min with (1) MES (0.1 M) (2.0 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 4.7, (2) HEPES (0.05 M) (1.2 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.3, (3) phosphate (0.1 M) (1.0 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.2, (4) Tris (0.05 M) (0.5 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.5, (5) CAPBSO (0.1 M) (2.0 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.5, (6) AMPBSO (0.1 M) (2.0 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.5, (7) MOPS (0.05 M) (0.5 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.5. (8) The solution contains SWCNT (0.5 mg/mL) in MOPS (0.1 M) (2.0 wt. %) with 0.05 M NaCl (0.15 wt. %) pH 7.5 buffer. All pictures are taken at initial dispersion state, before and after sonication for 30 min and then after sitting for 30 min.



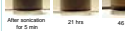
Ferritin Protein

- Iron storage protein in biological mechanisms in human, animal, and even bacteria.
- Contains up to ~4500 Fe³⁺ atoms.
- Stable and robust structure to withstand biological conditions of high temperature (up to 80 °C) and pH variations (2.0-10.0).
- Hydrophobic and hydrophilic channels.



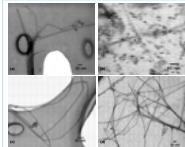
Core materials:

- Natural: Ferrihydrite, Fe₃(OH)₄(OH)
- Magnetic: magnetite (Fe₃O₄), Fe₂O₃
- Toxic: cerium(IV) oxide (CeO₂)
- Cu, Mn, Co, Cu₂S, Pt, Ru, Ni, W, etc.
- CdS, CdSe, CdTe, lanthanide doped QDs
- Prussian Blue



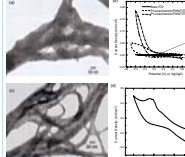
STEM image of chemically prepared Prussian Ferritins. STEM image was taken after addition 200 µL NaCl, into solution after 30 min and then sonication with NaCl for 10 min.

Photos of HEPES SWCNT-AMPBS dispersion containing 1.1 ml of cationized hole ferritin at 10 mg/mL. The dispersion solution was added to 0.1 M MOPS buffer without NaCl at pH 7.5 and HEPES SWCNT at 0.1 mg/mL. Total amount of SWCNT was approximately 7 µg. The ratio between the SWCNT and the Fe inside ferritin is 1 to 1.71 as a weight. The amount of Fe loading into the solution is 253.3 µg.



STEM images of ferritin protein interaction with FWCNTs in MOPS buffer. (a) hole ferritin and (b) cationized hole ferritin affinity for TWEEN 20 assisted FWCNTs suspension, and (c) hole ferritin and (d) cationized hole ferritin affinity for NaOBS assisted FWCNT suspension. The concentration of FWCNTs in water is 0.05 mg/mL and the ratio among the FWCNTs, the ferritin, and the surfactant is 1 : 4 : 100 times of weight.

STEM images of ferritin protein interaction with FWCNTs in MOPS buffer (0.1 M) with 0.05 M NaCl, pH 7.5. The ratio between the FWCNT and the hole ferritin are (a) 1 to 1 and (b) 1 to 4 by weight, respectively. (c) The image was taken with sample B after removal of unbound ferritins through filtering and washing. The ratio between ferritin and the cationized hole ferritin are (d) 1 to 1 and (e) 1 to 4 in terms of weight, respectively. (f) The image was taken with sample C after removal of unbound cationized ferritins through filtering and washing. The concentration of FWCNTs in water is 0.05 mg/mL.



(a) STEM image of Fe₃O₄ coated ferritin on FWCNTs. The ratio between the Fe₃O₄ and the Fe coated ferritin is 1 to 1.47 in terms of weight. The total Fe loading amount into the solution is 44.3 µg. (b) Cores: cationized ferritin of hole ferritin and Fe₃O₄ coated ferritin-Fe₃O₄ coated Fe₃O₄ electrode in 0.05 M phosphate buffer at pH 7.5 with buffer solution. Scan rate is 10 mV/s. (c) STEM image of Fe₃O₄ coated cationized ferritin on SWCNTs. The ratio between the SWCNT and the Fe₃O₄ coated ferritin is 1 to 1.4 in terms of weight. The total Fe loading amount into the solution is 33.3 µg. (d) Fe₃O₄ coated cationized ferritin-Fe₃O₄ coated Fe₃O₄ electrode in 0.05 M phosphate buffer at pH 7.5 with saturated solution. Scan rate is 10 mV/s.



Summary

We demonstrated high performance electrodes for oxygen reduction using CNTs conjugated with uniformly populated platinum nanoparticles generated by the reconstitution of ferritin proteins. These electrodes were achieved by effectively dispersing CNTs into the aqueous MOPS buffer containing P-coated cationized ferritins. The nanosized P-coated ferritins on the CNTs displayed good catalytic activity for the electrochemical reduction of oxygen which is applicable to biofuel cell and fuel cell applications.

Acknowledgement
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